

FORM PTO-1390 (Modified)  
(REV 11-98)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

## TRANSMITTAL LETTER TO THE UNITED STATES

002.00140

DESIGNATED/ELECTED OFFICE (DO/EO/US)

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

CONCERNING A FILING UNDER 35 U.S.C. 371

09/763848

INTERNATIONAL APPLICATION NO.

INTERNATIONAL FILING DATE

PRIORITY DATE CLAIMED

PCT/GB99/02761

19 August 1999

27 August 1998

## TITLE OF INVENTION

PROTEIN DOMAINS IN THE HEPATIC GLYCOGEN-TARGETTING SUBUNIT OF PROTEIN PHOSPHATASE  
1 AND METHODS OF MAKING AND USING THE SAME

## APPLICANT(S) FOR DO/EO/US

Patricia Townsend Wade Cohen; Christopher George Armstrong; Martin John Doherty

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
  - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☒ has been transmitted by the International Bureau.
  - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
  - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☐ have been transmitted by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
10. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

## Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☒ Certificate of Mailing by Express Mail
20. ☐ Other items or information:

U.S. APPLICATION NO. (IF KNOWN, SERIAL NUMBER) <b>09/763848</b>	INTERNATIONAL APPLICATION NO. <b>PCT/GB99/02761</b>	ATTORNEY'S DOCKET NUMBER <b>002.00140</b>
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21. The following fees are submitted:				<b>CALCULATIONS PTO USE ONLY</b>	
<b>BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :</b>					
<input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO ..... <b>\$1,000.00</b>					
<input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO ..... <b>\$860.00</b>					
<input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO ..... <b>\$710.00</b>					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) ..... <b>\$690.00</b>					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) ..... <b>\$100.00</b>					
<b>ENTER APPROPRIATE BASIC FEE AMOUNT =</b>				<b>\$860.00</b>	
Surcharge of <b>\$130.00</b> for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input checked="" type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).				<b>\$130.00</b>	
<b>CLAIMS</b>	<b>NUMBER FILED</b>	<b>NUMBER EXTRA</b>	<b>RATE</b>		
Total claims	13 - 20 =	0	x \$18.00	<b>\$0.00</b>	
Independent claims	4 - 3 =	1	x \$80.00	<b>\$80.00</b>	
Multiple Dependent Claims (check if applicable). <input type="checkbox"/>				<b>\$0.00</b>	
<b>TOTAL OF ABOVE CALCULATIONS =</b>				<b>\$1,070.00</b>	
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable). <input type="checkbox"/>				<b>\$0.00</b>	
<b>SUBTOTAL =</b>				<b>\$1,070.00</b>	
Processing fee of <b>\$130.00</b> for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).				<b>\$0.00</b>	
<b>TOTAL NATIONAL FEE =</b>				<b>\$1,070.00</b>	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input type="checkbox"/>				<b>\$0.00</b>	
<b>TOTAL FEES ENCLOSED =</b>				<b>\$1,070.00</b>	
				<b>Amount to be: refunded</b>	\$
				<b>charged</b>	\$

- ☒ A check in the amount of **\$1,070.00** to cover the above fees is enclosed.
- ☐ Please charge my Deposit Account No. \_\_\_\_\_ in the amount of \_\_\_\_\_ to cover the above fees.  
A duplicate copy of this sheet is enclosed.
- ☒ The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. **50-0772** A duplicate copy of this sheet is enclosed.

**NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.**

SEND ALL CORRESPONDENCE TO:

Susan J. Braman  
Braman & Rogalskyj, LLP  
P.O. Box 352  
Canandaigua, New York 14424-0352  
Tel 716-393-3002  
Fax 716-393-3001

Susan J. Braman  
SIGNATURE

Susan J. Braman  
NAME

34,103  
REGISTRATION NUMBER

27 February 2001  
DATE

002.00140

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s):	Cohen et al.	)	
		)	
Serial No.:	09/763,848	)	Examiner:
		)	
Filed:	September 26, 2001	)	
		)	Art Unit: 1646
For:	PROTEIN DOMAINS IN THE HEPATIC)		
	GLYCOGEN-TARGETTING SUBUNIT OF)		
	PROTEIN PHOSPHATASE 1 AND	)	
	METHODS OF MAKING AND USING	)	
	THE SAME	)	
		)	

PRELIMINARY AMENDMENT

Commissioner for Patents  
Box PCT  
U.S. Patent and Trademark Office  
Washington, D.C. 20231

Sir:

Please amend the subject application as follows:

After the Claims:

After the claims on page 37, add SEQUENCE LISTING pages 1-6 enclosed herewith.

REMARKS

Nucleotide Sequence Disclosure

In response to the notice of missing requirements, and in compliance with the sequence rules, applicants have enclosed the following:

1. A computer readable form (CRF) copy of a Sequence Listing in the form of a 3 1/2" diskette;
2. A paper copy of the Sequence Listing, pages 1-6; and
3. A statement that the content of the paper and computer readable form are the same and include no new matter.

Each of the sequences presented in the Sequence Listing were present in the application as originally filed. Accordingly no new matter is involved. Applicants respectfully request that the Sequence Listing be entered and maintain that the application now complies with the sequence rules.

Respectfully submitted,

May 3, 2002

Date

Susan J. Braman

Susan J. Braman

Reg. No. 34,103

Braman & Rogalskyj, LLP  
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Canandaigua, New York 14424-0352  
Tel: 585-393-3002  
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## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s): Cohen et al. )  
Serial No.: 09/763,848 ) Examiner:  
Filed: September 26, 2001 ) Art Unit: 1646  
For: PROTEIN DOMAINS IN THE HEPATIC )  
GLYCOGEN-TARGETTING SUBUNIT OF )  
PROTEIN PHOSPHATASE 1 AND )  
METHODS OF MAKING AND USING )  
THE SAME )

## STATEMENT IN ACCORDANCE WITH 37 C.F.R. §1.821(f)

Commissioner for Patents  
Box PCT  
U.S. Patent and Trademark Office  
Washington, D.C. 20231

Sir:

In accordance with 37 C.F.R. §1.821(f), this statement confirms that the contents of the Sequence Listing appearing on pages 1-6 (as submitted herewith) and on the computer readable 3.5" diskette (as submitted herewith) are the same and include no new matter.

Respectfully submitted,

May 3, 2002  
Date

Susan J. Braman  
Susan J. Braman  
Reg. No. 34,103

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Canandaigua, New York 14424-0352  
Tel: 585-393-3002  
Fax: 585-393-3001

# SEQUENCE LISTING

<110> Cohen, Patricia T.W.

Armstrong, Christopher G.

Doherty, Martin J.

<120> Protein Domains in the Hepatic Glycogen-Targetting  
Subunit of Protein Phosphatase 1 and Methods of Making  
and Using The Same

<130> 002.00140

<140> US 09/763,848

<141> 2001-09-26

<150> PCT/GB99/02761

<151> 1999-08-19

<150> GB 9818650.5

<151> 1998-08-27

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35 40 45

Lys Asp Thr Tyr Ala Gly Ser Asp Arg Asp Thr Phe Ser Phe Asp Ile  
50 55 60

Ser Leu Pro Glu Lys Ile Gln Ser Tyr Glu Arg Met Glu Phe Ala Val  
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Asp Leu Pro Pro Val Ile Pro Thr Glu Gln Lys Ile Glu Phe Cys Ile  
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20

25

30

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Gly Pro Ala Gly Pro Glu Gly Thr Glu Asp Val Phe Thr Phe Gly Phe  
 50 55 60

Pro Val Pro Pro Phe Leu Leu Glu Leu Gly Ser Arg Val His Phe Ala  
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&lt;211&gt; 102

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 9

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Arg Met Ser Leu Asp Asp Trp Gln Thr His Tyr Asp Ile Leu Ala Glu  
 35 40 45

Tyr Val Pro Asn Ser Cys Asp Gly Glu Thr Asp Gln Phe Ser Phe Lys  
 50 55 60

Ile Val Leu Val Pro Pro Tyr Gln Lys Asp Gly Ser Lys Val Glu Phe  
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&lt;211&gt; 117

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Ile Lys Phe Thr Phe Asn Ser Trp Arg Asp Ile His Tyr Val Thr Ala  
35 40 45

Asn Phe Asn Arg Thr Ile Asn Ser Asn Val Asp Glu Phe Lys Phe Thr  
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Ile Asp Leu Asn Ser Leu Lys Tyr Ile Leu Leu Ile Lys Arg Ile Ile  
65 70 75 80

Thr Met Glu Lys Asn Thr Ser Ser Cys Pro Leu Asn Ile Glu Leu Cys  
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35 40 45

Ser Tyr Ser Ala Pro Ile Ser Gly Ser Asn Tyr Glu Tyr Trp Thr Phe  
50 55 60

Ser Ala Ser Ile Asn Gly Ile Lys Glu Phe Tyr Ile Lys Tyr Glu Val

65

70

75

80

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&lt;211&gt; 40

&lt;212&gt; PRT

&lt;213&gt; Rabbit

&lt;400&gt; 12

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1 5 10 15

Val Ala Ala Ala Phe Pro Gly Asp Val Asp Arg Leu Arg Arg Met Ser  
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Leu Val Glu Glu Gly Ala Cys Lys  
35 40



## UNITED STATES PATENT AND TRADEMARK OFFICE

Commissioner for Patents, Box 101  
 United States Patent and Trademark Office  
 Washington, DC 20593-1010

U.S. APPLICATION NUMBER NO	FIRST NAMED APPLICANT	ATTY DOCKET NO
09/763,848	Patricia Townsend Wade Cohen	002.00140

INTERNATIONAL APPLICATION NO
------------------------------

PCT/GB99/02761

IA FILING DATE	PRIORITY DATE
08/19/1999	08/27/1998

Susan J Braman  
 Braman & Rogalskyj  
 PO Box 352  
 Canandaigua, NY 14424-0352

**CONFIRMATION NO. 3680**  
**371 FORMALITIES LETTER**  
**\*OC000000007156871\***  
 \*OC000000007156871\*

Date Mailed: 12/05/2001

### NOTIFICATION OF MISSING REQUIREMENTS UNDER 35 U.S.C. 371 IN THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US)

The following items have been submitted by the applicant or the IB to the United States Patent and Trademark Office as an Elected Office (37 CFR 1.495):

- U.S. Basic National Fees
- Priority Document
- Copy of IPE Report
- Copy of references cited in ISR
- Copy of the International Application
- Copy of the International Search Report
- Oath or Declaration
- Preliminary Amendments
- Request for Immediate Examination

The following items **MUST** be furnished within the period set forth below in order to complete the requirements for acceptance under 35 U.S.C. 371:

- The nucleotide and/or amino acid sequence disclosure contained in this application does not comply with the requirements for such a disclosure as set forth in 37 CFR 1.821-1.825 for the following reason(s):
  - This application does not contain a "Sequence Listing" as a separate part of the disclosure on paper copy or compact disc, as required by 37 CFR 1.821(c).
  - A copy of the "Sequence Listing" in computer readable form has not been submitted as required by 37 CFR 1.821(e).
  - APPLICANT MUST PROVIDE:
    - An initial or substitute computer readable form (CRF) of the "Sequence Listing."
    - An initial or substitute paper copy or compact disc of the "Sequence Listing," as well as an amendment directing its entry into the specification.
- For questions regarding compliance to 37 CFR 1.821-1.825 requirements, please contact:

- For Rules Interpretation, call (703) 308-4216
- To Purchase PatentIn Software, call (703) 306-2600
- For PatentIn Software Program Help, call (703) 306-4119 or e-mail at patin21help@uspto.gov or patin3help@uspto.gov

**ALL OF THE ITEMS SET FORTH ABOVE MUST BE SUBMITTED WITHIN TWO (2) MONTH FROM THE DATE OF THIS NOTICE OR BY 22 or 32 MONTHS (where 37 CFR 1.495 applies) FROM THE PRIORITY DATE FOR THE APPLICATION, WHICHEVER IS LATER. FAILURE TO PROPERLY RESPOND WILL RESULT IN ABANDONMENT.**

The time period set above may be extended by filing a petition and fee for extension of time under the provisions of 37 CFR 1.136(a).

Applicant is reminded that any communications to the United States Patent and Trademark Office must be mailed to the address given in the heading and include the U S application no shown above (37 CFR 1.5)

*A copy of this notice **MUST** be returned with the response.*

SHAKEEL AHMED

Telephone: (703) 305-3659

**PART 1 - ATTORNEY/APPLICANT COPY**

U.S. APPLICATION NUMBER NO.	INTERNATIONAL APPLICATION NO	ATTY DOCKET NO
09/763,848	PCT/GB99/02761	002.00140

002.00140

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s): Cohen et al. )  
Serial No.: To be assigned (U.S. National )  
Stage of PCT/GB99/02761) )  
Filed: Herewith )  
For: PROTEIN DOMAINS IN THE HEPATIC GLYCOGEN- )  
TARGETTING SUBUNIT OF PROTEIN )  
PHOSPHATASE 1 AND METHODS OF MAKING )  
AND USING THE SAME )

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents  
Washington, D.C. 20231  
Box PCT

Dear Sir:

Please amend the subject application as follows  
(amendments refer to the claims as published):

In the Claims:

Please amend claims 12 and 13 as follows:

12. (Amended) A compound which is identifiable by the  
method of claim 9 [any one of Claims 9 to 11].

13. (Amended) A method of reducing the blood glucose  
level of a mammalian animal comprising administering a  
therapeutically effective amount of [an inhibitor] a compound  
[as defined in any one of Claims 1 to 12] which is capable of  
blocking the interaction of phosphorylase a with the glycogen-  
targeting subunit (G<sub>r</sub>) of protein phosphatase 1.

09/763848-00001

**REMARKS**

Claims 1-14 are presented for examination in the subject application, as published. By this preliminary amendment, claims 12 and 13 have been amended.

Respectfully submitted,

Dated: 27 February 2001

Susan J. Braman  
Susan J. Braman  
Reg. No. 34,103

Braman & Rogalskyj, LLP  
P.O. Box 352  
Canandaigua, New York 14424-0352  
Telephone: (716) 393-3002  
Facsimile: (716) 393-3001

0976349-09604  
T09260-8426260

PROTEIN DOMAINS IN THE HEPATIC GLYCOGEN-TARGETTING SUBUNIT OF PROTEIN PHOSPHATASE 1 AND METHODS OF MAKING AND USING THE SAME

The present invention relates to compounds useful in the treatment of disorders associated with abnormal blood glucose levels, particularly in the prevention of phosphorylase-a binding to the glycogen targeting subunit ( $G_L$ ) of protein phosphatase 1 (PP1). Such compounds are useful for increasing glycogen synthesis and thereby reducing blood glucose levels. The compounds find utility in the treatment of disorders, such as type I and type II diabetes, associated with higher than normal levels of blood glucose (hyperglycaemia).

Most of the adverse physiological consequences in type I and type II diabetes arise from the higher than normal levels of blood glucose. Although high blood glucose levels can be reduced by administration of insulin in type 1 diabetes and by dietary restrictions in the case of type II diabetes, a drug which aids reduction of blood glucose levels would be advantageous in the treatment of these disorders. The liver, which is the main organ regulating glucose homeostasis, is able to store glucose in the form of glycogen and the synthesis of hepatic glycogen from glucose is under the control of hepatic glycogen synthase.

Protein phosphatase 1 is major protein serine/threonine phosphatase in eukaryotic cells, which regulates numerous distinct cellular processes. This is achieved by the interaction of the catalytic subunit of PP1 with a diverse range of targeting subunits that localise PP1C to specific sites within the cell, modulate its activity towards particular substrates and allow its activity to respond to extracellular signals [1,2].

The family of proteins that target PP1 to glycogen and regulate its activity towards the enzymes of glycogen metabolism consists of four members,



$G_M$ /PPP1R3,  $G_L$ /PPP1R4, PPP1R5 and PPP1R6/PTG [3-8]. The liver specific glycogen-targeting subunit,  $G_L$  is a 33 kDa protein [5,9] that, when bound to PP1, enhances the rate at which the latter dephosphorylates and activates the rate determining enzyme in glycogen synthesis, glycogen synthase, whilst suppressing the rate at which it inactivates glycogen phosphorylase. The stimulation of hepatic glycogenolysis by glucagon (acting via cyclic AMP and PKA) and  $\alpha$ -adrenergic agonists (acting via  $Ca^{2+}$ ) is achieved by activation of phosphorylase kinase respectively, which increases the levels of the active phosphorylated form of glycogen phosphorylase (phosphorylase  $a$ ). In addition, phosphorylase  $a$  binds to  $G_L$  and potently inhibits its glycogen synthase phosphatase activity thereby inhibiting glycogen synthesis. Insulin lowers hepatic cAMP levels, causing a reduction in the level of phosphorylase  $a$  and alleviation of the phosphorylase  $a$  -mediated inhibition of the PP1 $G_L$  complex, while the binding of glucose to phosphorylase  $a$ , increases the rate at which phosphorylase is inactivated. These mechanisms contribute to the stimulation of glycogen synthesis by insulin and high blood glucose [10]. The inhibition of the PP1 $G_L$  complex by phosphorylase  $a$  occurs at nanomolar concentrations and is thought to be via an allosteric mechanism since the  $K_m$  for phosphorylase  $a$  as a substrate is in the micromolar range [11]. This view is strengthened by the finding that phosphorylase  $a$  (but not phosphorylase  $b$ ) binds directly to  $G_L$  in protein blotting experiments [5,9].

Recent studies identified conserved regions between the glycogen targeting subunits  $G_M$ /PPP1R3 and  $G_L$ /PPP1R4 [5]. A peptide corresponding to one of these regions,  $G_M$ 63-75 was shown to bind PP1 and the amino terminal 38 residues of the myofibrillar targeting subunit of PP1 were also

demonstrated to interact with PP1 [12]. The G<sub>M</sub> 63-75 peptide, which contains a small motif common to the myofibrillar binding subunit and many other of the PP1 targeting subunits has been crystallised as a complex with PP1 and its structure solved to 2.8 Å resolution [2]. This motif, Lys/Arg-Val/Ile-Xaa-Phe/Trp, which has also been identified by a random peptide library approach [13], is found in all the four glycogen targeting subunit and is located at residues 60-64 of G<sub>L</sub>. However, incubation of the PP1G<sub>L</sub> complex purified from hepatic glycogen-protein particles with a PP1-binding peptide from G<sub>M</sub> failed to dissociate the PP1-G<sub>L</sub> complex [12], even though the peptide abolished the suppression of phosphorylase phosphatase activity conferred on PP1 by association with G<sub>L</sub>.

The present invention seeks to provide biological materials and methods which may be useful in the treatment of disorders, especially those such as diabetes type I and type II, associated with higher than normal levels of blood glucose.

According to a first aspect of the invention there is provided the use in medicine of a compound which is capable of blocking the interaction of phosphorylase *a* with the glycogen-targeting subunit (G<sub>L</sub>) of protein phosphatase 1.

Preferably, the compound is for use in the manufacture of a medicament for use in treating disorders associated with higher than normal levels of blood glucose. Preferably the medicament is for use in the treatment of a disorder selected from type I and/or type II diabetes.

Preferably, the compound is a polypeptide comprising the sequence of the C-terminal 16 amino acids of human  $G_L$  sequence, or a fragment or variant thereof of which is capable of binding phosphorylase  $\alpha$ .

Thus, the sequence may be PEWPSYLGYEKLGOPY, which may be the sequence of the C-terminal 16 amino acids of rat liver  $G_L$ .

By "variant" we include the meaning of polypeptides comprising an amino acid sequence which, although not identical to the 16 amino acid sequence, are capable of binding phosphorylase  $\alpha$ .

By "fragment" we include the meaning that the polypeptide comprises less than the 16 amino acid sequence mentioned above, but is capable of binding phosphorylase  $\alpha$ .

The identification of variants and fragments within the scope of the invention can be carried out using the methods described herein.

Preferably the polypeptide increases the activity of hepatic glycogen synthase.

Polypeptides in which one or more of the amino acid residues are chemically modified, before or after the polypeptide peptide is synthesised, may be used in accordance with the invention, providing that the function of the peptide, namely the blocking of the interaction between  $G_L$  and phosphorylase  $\alpha$ , remains substantially unchanged. Such modifications include forming salts with acids or bases, especially physiologically acceptable organic or inorganic acids and bases, forming an ester or amide of a terminal carboxyl group, and attaching amino acid

protecting groups such as N-t-butoxycarbonyl. Such modifications may protect the peptide from *in vivo* metabolism.

The peptides may be present as single copies or as multiples, for example tandem repeats. Such tandem or multiple repeats may increase the activity of the polypeptide in blocking the binding of  $G_L$  and phosphorylase  $\alpha$ .

In a second aspect, the invention provides a pharmaceutical composition comprising an inhibitor compound which is capable of blocking the interaction of phosphorylase  $\alpha$  with the glycogen targeting subunit ( $G_L$ ) of protein phosphatase (PP1), together with a pharmaceutically acceptable excipient or carrier. Preferably the inhibitor compound comprises a polypeptide having the sequence of the C-terminal 16 amino acids of human  $G_L$  sequence, or a fragment or variant thereof of which is capable of binding phosphorylase  $\alpha$ , for example the 16 amino acid sequence PEWPSYLGYEKLGPIYY or a fragment or variant thereof of which is capable of binding phosphorylase  $\alpha$ .

In a third aspect, the invention provides a method of identifying an inhibitor compound that is capable of blocking the interaction of phosphorylase  $\alpha$  with the glycogen-targeting subunit of PP1 comprising: providing a polypeptide comprising the sequence of the C-terminal 16 amino acids of human  $G_L$  sequence, or a fragment or variant thereof of which is capable of binding phosphorylase  $\alpha$ , for example the 16 amino acid sequence PEWPSYLGYEKLGPIYY, or a fragment or variant thereof which binds phosphorylase  $\alpha$ ; providing a test compound; and

comparing the binding of the polypeptide by phosphorylase  $\alpha$  in the presence or absence of the test compound; an inhibitor compound being identified by reduced binding in the presence of the test compound.

The inhibitor compound may be a drug-like compound or lead compound for the development of a drug-like compound. Thus, the method may be a method for identifying a drug-like compound or lead compound for the development of a drug-like compound that is capable of blocking the interaction of phosphorylase  $\alpha$  with the glycogen-targeting subunit of PP1.

The term "drug-like compound" is well known to those skilled in the art, and may include the meaning of a compound that has characteristics that may make it suitable for use in medicine, for example as the active ingredient in a medicament. Thus, for example, a drug-like compound may be a molecule that may be synthesised by the techniques of organic chemistry, less preferably by techniques of molecular biology or biochemistry, and is preferably a small molecule, which may be of less than 5000 daltons molecular weight and which may be water-soluble. A drug-like compound may additionally exhibit features of selective interaction with a particular protein or proteins and be bioavailable and/or able to penetrate target cellular membranes, but it will be appreciated that these features are not essential.

The term "lead compound" is similarly well known to those skilled in the art, and may include the meaning that the compound, whilst not itself suitable for use as a drug (for example because it is only weakly potent against its intended target, non-selective in its action, unstable, poorly soluble, difficult to synthesise or has poor bioavailability) may provide a

starting-point for the design of other compounds that may have more desirable characteristics.

The compounds identified in the method may themselves be useful as a drug or they may represent lead compounds for the design and synthesis of more efficacious compounds.

The inhibitor compound may be an antibody or immunoglobulin-type molecule or a fragment thereof, as well known to those skilled in the art. An antibody or immunoglobulin-type molecule or a fragment thereof that cross-reacts/binds with a polypeptide having the sequence of the C-terminal 16 amino acids of human  $G_L$  sequence, or a fragment or variant thereof of which is capable of binding phosphorylase  $a$ , for example the 16 amino acid sequence PEWPSYLGYEKLGPIYY or a fragment or variant thereof of which is capable of binding phosphorylase  $a$  may be such an inhibitor compound. An antibody or immunoglobulin-type molecule or a fragment thereof that cross-reacts/binds to  $G_L$  may also be such an inhibitor compound. Such antibodies or immunoglobulin-type molecules or fragments thereof may be prepared by methods well known to those skilled in the art.

It will be appreciated that by "phosphorylase  $a$ " is included variants, fragments and fusions of phosphorylase  $a$  that have interactions or activities which are substantially the same as those of phosphorylase  $a$  with  $G_L$  but which may be more convenient to use in an assay. For example, a fusion of phosphorylase  $a$  may be useful since said fusion may contain a moiety which may allow the fusion to be purified readily. It is preferred that the phosphorylase  $a$  is phosphorylase  $a$  from rabbit skeletal

muscle. Phosphorylase *a* may be obtained from Sigma, Sgima-Aldrich Company Ltd, Fancy Road, Poole, Dorset, BH12 4QH.

Preferably, the phosphorylase *a* is labelled and the binding of phosphorylase *a* to the polypeptide is determined by measuring the amount of label quantitatively or qualitatively.

Conveniently, the phosphorylase *a* is labelled with a label selected from digoxigenin,  $^{33}\text{P}$  and  $^{32}\text{P}$ . Phosphorylase *a* labelled with  $^{32}\text{P}$  or  $^{33}\text{P}$  may be obtained by phosphorylation by phosphorylase kinase, as described in Cohen *et al* (1988) *Meth Enzymol* **159**, 399-408. Phosphorylase kinase may be obtained from Sigma.

The disruption of the interaction between the said polypeptide and phosphorylase *a* can be measured *in vitro* using methods well known in the art of biochemistry and include any methods which can be used to assess protein-protein interactions.

The said interaction can also be measured within a cell, for example using the yeast two hybrid system as is well known in the art.

It will be appreciated that screening assays which are capable of high throughput operation will be particularly preferred. Examples may include cell based assays and protein-protein binding assays. An SPA-based (Scintillation Proximity Assay; Amersham International) system may be used. For example, beads comprising scintillant and the polypeptide having the sequence of the C-terminal 16 amino acids of human  $G_L$  sequence, or a fragment or variant thereof of which is capable

of binding phosphorylase *a*, for example the 16 amino acid sequence PEWPSYLGYEKLGPIYY or a fragment or variant thereof of which is capable of binding phosphorylase *a* may be prepared. The beads may be mixed with a sample comprising  $^{32}\text{P}$ - or  $^{33}\text{P}$ -labelled phosphorylase *a* and with the test compound. Conveniently this is done in a 96-well format. The plate is then counted using a suitable scintillation counter, using known parameters for  $^{32}\text{P}$  SPA assays. Only  $^{32}\text{P}$  that is in proximity to the scintillant, i.e. only that bound to the polypeptide, is detected. Variants of such an assay, for example in which the polypeptide is immobilised on the scintillant beads *via* binding to an antibody, may also be used.

Other methods of detecting polypeptide/polypeptide interactions include ultrafiltration with ion spray mass spectroscopy/HPLC methods or other physical and analytical methods. Fluorescence Energy Resonance Transfer (FRET) methods, for example, well known to those skilled in the art, may be used, in which binding of two fluorescent labelled entities may be measured by measuring the interaction of the fluorescent labels when in close proximity to each other.

In a fourth aspect, the invention provides a compound which is identifiable by the methods according to the third aspect of the invention.

In a fifth aspect, the invention provides a method of reducing the blood glucose level of a mammalian animal comprising administering a therapeutically effective amount of an inhibitor compound as defined in accordance with the previous aspects of the invention. Preferably the mammalian animal is a human.



By "therapeutically effective amount" we include the meaning that enough of the compound is administered to produce a beneficial effect in the recipient, for example a beneficial decrease in hyperglycaemia.

A sixth aspect of the invention provides a method of identifying a compound which mimics the effect of phosphorylase  $\alpha$  on  $G_L$ , the method comprising contacting said compound with  $G_L$  and determining whether, in the presence of the compound,  $G_L$  adopts the properties of  $G_L$  in the presence of phosphorylase  $\alpha$ .

By "mimics the effect of phosphorylase  $\alpha$ " we include the meaning that the compound modifies a property of  $G_L$  in such a way that  $G_L$  acts, in at least one respect, like  $G_L$  that is interacting with phosphorylase  $\alpha$ .

It will be appreciated that the  $G_L$  may be bound to PP1c, ie may be in the form of PP1 $G_L$  and that the effect of the compound may be assessed by measuring the activity of PP1 $G_L$ , as well known to those skilled in the art, for example as described in WO97/37224 and references therein. Thus, the dephosphorylation of glycogen synthase by PP1 $G_L$  may be measured. Thus a compound may be selected that decreases the activity of PP1 $G_L$ . It will be appreciated that the method may include the conduct of a screen or screens to determine that the compound interacts with the  $G_L$  subunit and not with the PP1c subunit.

The said compound may be a drug-like compound or lead compound for the development of a drug-like compound. Thus, the method may be a method for identifying a drug-like compound or lead compound for the

development of a drug-like compound that is capable of mimicking the effect of phosphorylase  $\alpha$  on  $G_L$  (a mimic compound).

In a seventh aspect, the invention provides a compound which is identifiable by the methods according to the sixth aspect of the invention.

In an eighth aspect, the invention provides a method of increasing the blood glucose level of a mammalian animal comprising administering a therapeutically effective amount of an inhibitor compound as defined in accordance with the previous aspects of the invention. Preferably the mammalian animal is a human. The mammalian animal, for example human, may have hypoglycaemia.

By "therapeutically effective amount" we include the meaning that enough of the compound is administered to produce a beneficial effect in the recipient, for example a beneficial decrease in hypoglycaemia.

A further aspect of the invention provides a kit of parts useful in carrying out the method of the third aspect of the invention. Thus such a kit may comprise phosphorylase  $\alpha$  and a polypeptide comprising the sequence of the C-terminal 16 amino acids of human  $G_L$  sequence, or a fragment or variant thereof of which is capable of binding phosphorylase  $\alpha$ , for example the 16 amino acid sequence PEWPSYLGYEKLGPYY, or a fragment or variant thereof which binds phosphorylase  $\alpha$ .

Preferred embodiments of the invention will now be described by way of example, with reference to the accompanying figures.

Figure 1. Schematic representation of the  $G_L$  and truncated forms generated by PCR or restriction digestion. The ability of  $G_L$  and the  $G_L$  fragments to bind PP1, glycogen and phosphorylase  $\alpha$  is indicated. + indicates that binding was observed; - denotes that binding was tested but not detected; a indicates that aggregation of  $G_L$  and its derivatives precluded testing of glycogen binding; interactions that were not tested are left blank.

Figure 2. Identification of the  $G_L$  region required for interaction with glycogen. GST- $G_L$  fusion proteins containing truncated  $G_L$  coding regions were examined for their ability to co-sediment with glycogen as described hereinafter. The supernatant (S) and the pellet (P) fractions obtained in the absence and presence of glycogen were subjected to SDS/PAGE on 12.5% polyacrylamide gels, transferred to nitrocellulose membranes and immunoblotted with affinity purified anti- $G_L$  protein antibodies. The position of the standard marker proteins, glycogen phosphorylase (97kDa), bovine serum albumin (66kDa), ovalbumin (43kDa), and carbonic anhydrase (30kDa) are indicated.

Figure 3. Identification of the binding site for phosphorylase  $\alpha$  lies in the 16 carboxy-terminal amino acids of  $G_L$ . GST- $G_L$  (1-284) and GST-fusions containing truncated  $G_L$  coding regions (2 $\mu$ g) were separated on 12.5% SDS-polyacrylamide gels and either (A) stained with Coomassie blue or (B) transferred to nitrocellulose and probed with 100 nM  $^{32}$ P-labelled phosphorylase  $\alpha$ . The standard marker proteins are as in Fig. 2.

Figure 4. Identification of the region binding PP1 in  $G_L$ . GST- $G_L$  (1-284) and GST-fusions containing truncated  $G_L$  coding regions (2 $\mu$ g) were

separated on 12.5% SDS-polyacrylamide gels and either (A) stained with Coomassie blue or (B) transferred to nitrocellulose and probed with Digoxigenin-labelled PP1 $\gamma$ . The standard marker proteins are as in Fig. 2.

Figure 5. Comparison of the sequences of rat M<sub>110</sub> 1-38 peptide and rabbit G<sub>M</sub> 63-93 peptide with G<sub>L</sub> in the region of the RVSF motif (underlined). Three basic residues (Lys or Arg) preceding the RVSF motif that are conserved in M<sub>110</sub> and G<sub>L</sub> but not present in the G<sub>M</sub> 63-93 peptide are double underlined. No other clear sequence similarities between M<sub>110</sub> 1-38 peptide and G<sub>L</sub> were apparent in the region preceding that shown.

Figure 6. Comparison of the polysaccharide binding domain of the mammalian glycogen targeting subunits, *S. cerevisiae* GAC1, *Rhizopus oryzae* glucoamylase (AMYL) with phosphorylase (PHOS) glycogen binding (storage) site. The sequences are rat G<sub>L</sub>[5], human PPP1R5[6], human PPP1R6[7], human G<sub>M</sub>[4], *S.cerevisiae* GAC1[19], *R. oryzae* glucoamylase [20] and rabbit skeletal muscle glycogen phosphorylase [21]. A consensus sequence for the polysaccharide binding domain of the glycogen targeting subunits and glucoamylase is shown. Conserved residues are underlined and identical residues are double underlined. The maltohepatose binding residues in phosphorylase are marked with an asterisk.

Figure 7. Schematic representation of the domains in G<sub>L</sub> that interact with PP1, glycogen and phosphorylase  $\alpha$ . PP1 binds to the Arg-Val-Ser-Phe motif located at residues 61-64. The basic sequence preceding this

motif is also likely to be involved in binding PP1. Residues 134-231 include the conserved residues among the glycogen subunits that lie in the 94-257 fragment of  $G_L$  that co-sediments with glycogen. Phosphorylase  $\alpha$  binds to the carboxyterminal terminal 16 amino acids of  $G_L$ . The relevant amino acid positions in  $G_L$  are indicated above.

## MATERIALS AND METHODS

### 1. *Production of glutathione S-transferase- $G_L$ fusion proteins*

The pGEX- $G_L$  construct containing the complete coding region of  $G_L$  [5] was used as a template in polymerase chain reactions using primer pairs to generate a variety of  $G_L$  coding region fragments (Fig. 1). The 5' coding primers all contained an *NdeI* site and the 3' reverse primers contained the termination codon and a *XhoI* site as described in [5]. The PCR products were then subcloned into the TOPO 2.1 PCR cloning vector (Invitrogen, Leek, The Netherlands) and verified by sequencing on an Applied Biosystems 373A automated DNA sequencer using Taq dye terminator cycle sequencing. The  $G_L$  coding region fragments were subsequently excised by restriction cleavage with *NdeI* and *XhoI* and ligated into the pGEX-AH vector digested with the same restriction enzymes. The  $G_L$  1-94 and 1-170 truncations were generated by excision of a *SacI-SacI* and a *HindIII-HindIII* restriction fragment respectively from the pGEX- $G_L$  construct followed by religation of the plasmid. The  $G_L$  94-170 truncation was generated by *NdeI-HindIII* digestion of pGEX- $G_L$  94-257 followed by ligation of the fragment into pGEX-AH vector digested with the same enzymes. Site directed mutation of the single mutants N152A, K157A, the double mutant N152A + K157A and the triple mutant K149A +

N152A + L153A were generated as described in [14] using pGEX-G<sub>L</sub> 94-257 as template. The resultant constructs encoded glutathione S-transferase (GST) fused to G<sub>L</sub> coding region fragments of various lengths. Soluble GST-G<sub>L</sub> fusion-proteins were obtained by growing *E. coli*, transformed with the various pGEX-G<sub>L</sub> deletion constructs, in LB medium containing 100 µg/ml ampicillin and inducing expression in log phase growth at an A<sub>600nm</sub> of 0.5 with 0.2 mM isopropyl-thio-β-D-galactopyranoside. After 16 hours further growth at 26°C-28°C, the *E. coli* were harvested and soluble GST-G<sub>L</sub> fusion proteins were purified in glutathione agarose as described in [5].

## 2. *Interaction of GST-G<sub>L</sub> fusion proteins with digoxigenin-PPl<sub>y</sub> and phosphorylase a*

GST-G<sub>L</sub> fusion proteins were separated on SDS-PAGE and transferred to nitrocellulose membranes. They were probed with digoxigenin-PPl<sub>y</sub> as described [7]. Alternatively they were examined for binding of <sup>32</sup>P-labelled phosphorylase *a*. Non-specific binding to the membranes was blocked by incubation in 5% (w/v) marvel dried milk powder, 25 mM Tris/HCl pH 7.5, 500 mM NaCl for 16 hours. The samples were then probed for 3 h with <sup>32</sup>P-labelled phosphorylase *a* (100 nM) in 25 mM Tris/HCl pH 7.5, 250 mM NaCl, 1 mg/ml bovine serum albumin. The membranes were subsequently washed (3 x 30 min) with 25 mM Tris/HCl pH 7.5 before autoradiography.

### 3. *Co-sedimentation of GST-G<sub>L</sub> fusion proteins with hepatic glycogen*

Protein free glycogen was prepared by the following protocol. Glycogen-protein particles were isolated from the livers of New Zealand White rabbits [15]. Protein was then stripped from the glycogen by boiling for 5 min in 1 % (w/v) sodium dodecyl sulphate (SDS). The suspension was cooled to room temperature and centrifuged for 60 min at 100,000 x g. The 100,000 x g pellet was then resuspended in water, and the centrifugation and resuspension procedure repeated a further two times in order to completely remove residual SDS and protein. Any contaminating nucleotides were removed by incubating the glycogen for 15 min with mixed bed resin, AG 501-X8(D). The resin was removed by filtration and the glycogen concentration determined by the phenol/sulphuric acid method [16].

Protein free glycogen (10 mg/ml) in 50 mM Tris/HCl pH 7.5, 150 mM NaCl, 0.1 % (v/v) 2-mercaptoethanol, 0.02% (w/v) Brij-35, 0.1 mg/ml bovine serum albumin was mixed with GST-GL fusion proteins (50 nM). After incubation on ice for 30 min, the samples were centrifuged for 90 min at 100,000 x g. The supernatant and pellet fractions were denatured in SDS, subjected to SDS-polyacrylamide gel electrophoresis and transferred to nitrocellulose. The membranes were incubated overnight in 25 mM Tris/HCl pH 7.5, 250 mM NaCl, 0.1 % (w/v) Tween-20, 10% dried milk powder before probing with affinity purified sheep anti-GL protein antibodies (100 ng/ml in 25 mM Tris/HCl pH 7.5, 250 mM NaCl, 0.1 % (w/v) Tween-20, 3% (w/v) dried milk powder), followed by several washes in the same buffer (without the dried milk powder) and incubation with horse radish peroxidase-conjugated anti-sheep antibodies (Pierce,

UK). Immunoreactive bands were visualised using the enhanced chemiluminescence system (Amersham International, Bucks, UK.)

#### 4. Results

##### 4.1 Residues 94-257 of $G_L$ are required for binding to glycogen

GST- $G_L$ , containing the full-length  $G_L$  coding region, and several of the GST- $G_L$  truncations {GST- $G_L$ (94-216), GST- $G_L$ (94-170), GST- $G_L$ (134-170), GST- $G_L$ (134-216), GST- $G_L$ (134-257)} exhibited a strong tendency to aggregate and were pelleted at 100 000 x g for 1 h, even in the absence of glycogen. Therefore these constructs could not be tested for glycogen dependent sedimentation. Of the GST-fusion proteins that did not aggregate, GST- $G_L$ (94-284) and GST- $G_L$ (94-257) were both detected exclusively in the 100,000 x g pellet obtained by centrifugation in the presence of glycogen (Fig. 2). In contrast GST- $G_L$ (170-216) and GST- $G_L$ (170-257) did not bind to glycogen, being detected exclusively in the 100,000 x g supernatant fraction in the presence of glycogen (Fig. 2). GST- $G_L$ (94-257) carrying the single mutations N152A or K157A, the double mutation N152A + K157A, or the triple mutation K149A + N152A + L153A were all found to sediment in the presence of glycogen (data not shown).

##### 4.2 The phosphorylase $\alpha$ binding site lies in the C-terminal 16 amino acids of $G_L$

In order to identify the region of  $G_L$  that is responsible for the binding of phosphorylase  $\alpha$ , GST- $G_L$  and its truncated forms were transferred to



nitrocellulose membranes and tested for their ability to bind  $^{32}\text{P}$ -phosphorylase  $\alpha$ .  $^{32}\text{P}$ -labelled phosphorylase  $\alpha$  was found to bind to GST- $G_L$  containing the entire  $G_L$  coding region and to GST- $G_L(216-284)$ , GST- $G_L(257-284)$  and GST- $G_L(269-284)$  but not to GST- $G_L(1-216)$ , GST- $G_L(1-257)$  and GST- $G_L(1-271)$  {Fig. 3 and data not shown}. These results indicate that the phosphorylase  $\alpha$  binding domain lies in the carboxy-terminal 16 amino acids of  $G_L$ .

#### 4.3 *The PPI-binding domain lies between residues 59 and 94 of $G_L$*

The GST- $G_L$  truncations were also tested for their ability to bind digoxigenin-labelled PP1 $\gamma$  after transfer to nitrocellulose membranes. Fig. 4 shows that digoxigenin-PP1 binds to GST- $G_L(1-284)$ , GST- $G_L(1-94)$  and GST- $G_L(59-284)$  but not to GST- $G_L(1-59)$  or GST- $G_L(94-284)$ , GST- $G_L(134-284)$  or GST- $G_L(170-284)$ . From these interactions, the principal PP1 binding domain must lie between residues 59 and 94 of  $G_L$ . Several proteolytically degraded fragments present in the preparations were also recognised by digoxigenin-PP1, in particular a minor 35 kDa Coomassie-blue staining band that migrated slightly faster than GST- $G_L(1-94)$  but slower than GST- $G_L(1-59)$ . Since this proteolytic fragment was retained on glutathione-Sepharose, it is likely to comprise GST linked to the first 75-80 residues of  $G_L$ . The strong signal with digoxigenin-PP1 may be explained by a more effective renaturation of this fragment from SDS on the nitrocellulose membrane.

## 5. Discussion

Here we have identified three distinct functional domains on the rat liver glycogen-targeting subunit of protein phosphatase 1. The section comprising amino acids 59-94 is both necessary and sufficient for binding to PP1. This region contains the sequence, Arg-Val-Ser-Phe which conforms to the consensus PP1-binding motif determined for other PPI-binding subunits. The data provides further evidence for the importance of this short motif in the binding of PP1 to its targeting subunits. The results also indicate that no other domains outside of residues 59-94 can initiate and maintain an interaction with PP1 independently of the RVSF motif. Previous experiments [12] demonstrated that a peptide comprising the amino terminal 38 residues of the  $M_{110}$  subunit of the myosin targeting subunit of PP1 ( $M_{110}$  1-38) could disrupt the PP1 $G_L$  complex, while a peptide comprising residues 63-93 of the skeletal muscle glycogen targeting subunit ( $G_M$  63-93) did not abolish binding of  $G_L$  to PP1. This data suggests that secondary sites of PP1 $G_L$  interaction are likely to involve residues that are identical in  $M_{110}$  1-38 and  $G_L$  but distinct (or not included) in  $G_M$  63-93. A comparison of the sequences of  $M_{110}$  1-38 peptide and  $G_M$  63-93 peptide with  $G_L$  in the region of the RVSF motif identifies 3 basic residues (Lys or Arg) preceding the RVSF motif that are identical in  $M_{110}$  and  $G_L$  but are not present in the  $G_M$  63-93 peptide (Fig. 5). The basic residues in positions -2, -4 and -5 with respect to the RVSF motif may therefore provide the secondary interactions of  $G_L$  with PP1 that are not disrupted by the  $G_M$  63-93 peptide. The crystal structure of PP1 complexed to the  $G_M$  63-75 peptide reveals the presence of an acidic domain in PP1, that ties to the amino terminal end of the RVSF motif in

the bound peptide and thus has the potential to interact with the basic residues in  $G_L$  and the  $M_{110}$  peptide [2].

The sequence similarity noted previously between the mammalian glycogen targeting subunits and glucoamylase from *Rhizopus oryzae*, which binds starch, spans a region comprising amino acids 134-231 of  $G_L$  [5,6] and (Fig. 6). This present study demonstrates that the region 94-257 of  $G_L$  is capable of binding to glycogen, while the truncated GST fusion proteins GST- $G_L$ (170-216) or GST- $G_L$ (170-257) fail to bind to glycogen. The interaction of GST- $G_L$ (94-170) or GST- $G_L$ (134-170) with glycogen could not be tested due to the aggregation of this fragment. However, residues 148-168 of  $G_L$  show some sequence similarities (Fig. 6) to the region in phosphorylase identified in crystallographic studies to bind to maltoheptaose and believed to bind to glycogen *in vivo* [17, 181]. The crystal structure of phosphorylase shows that the side chains of conserved hydrophobic amino acids in this section point inwards and appear to be involved in maintaining the orientation of the  $\alpha$ -helix that binds maltoheptaose. Of the residues shown to bind to maltoheptaose, only the Asn corresponding to Asn152 in  $G_L$  is identical in the glycogen targeting subunits and phosphorylase. However, mutation of N152A in GST- $G_L$ (94-257) did not prevent this fragment binding to glycogen, nor did the double mutation of K157A+N152A. The triple mutation K149A + N152A + L153A of residues which are conserved in the glycogen targeting subunits and are aligned with those binding maltoheptaose in phosphorylase (Fig. 6) also did not prevent the binding of GST- $G_L$ (94-257) to glycogen. The results suggest that all of the conserved sections in  $G_L$  (134-231) are likely to be required for binding to glycogen either by making direct contact with glycogen or by contributing to structural elements required for this binding. They further indicate that the binding

site of the PP1 glycogen targeting subunit and glucoamylase to polysaccharides is distinct from the glycogen binding (storage) site of phosphorylase.

The allosteric inhibition of the glycogen synthase phosphatase activity of PP1GL by phosphorylase *a* is mediated by the binding of phosphorylase *a* to G<sub>L</sub> [5,9,11]. The results presented here demonstrate that the 16 amino acids at the C-terminus of G<sub>L</sub> are essential for the interaction with phosphorylase *a* (Fig. 7). However, although this short region is sufficient for the binding of phosphorylase *a*, it is likely that other regions of G<sub>L</sub> are required to transmit the allosteric effect of this molecule to the active site of PP1. Nevertheless, the unexpected identification of a short sequence in G<sub>L</sub> as crucial for the binding of phosphorylase *a* and therefore also for inhibition of glycogen synthase phosphatase activity provides a rationale for searching for small molecules that might block this inhibition. Raising the level of glycogen synthase phosphatase and consequently glycogen synthase may be useful in disorders, such as diabetes, where hyperglycaemia is a severe problem.

The other three glycogen-binding subunits, G<sub>M</sub>/PPP1R3, PPP1R5 and PPP1R6 do not show significant sequence similarity to the carboxy-terminus of G<sub>L</sub> [7] which explains why G<sub>M</sub> is not inhibited by phosphorylase *a* [10] and PPP1R5 and PPP1R6 have not been found to bind phosphorylase *a* after transfer to nitrocellulose membranes [5, 7]. PTG, the mouse homologue of the human glycogen-targeting subunit PPP1R5, has been reported to bind to phosphorylase *a*, as well as to several other regulatory enzymes of glycogen metabolism [8]. If this is the case, then the amino acid sequence binding, phosphorylase *a* in PTG is significantly different from that in G<sub>L</sub>.

6. *Method of identifying an inhibitor compound which is capable of blocking the interaction of phosphorylase  $\alpha$  with the glycogen-targeting subunit of PP1.*

GST-G<sub>L</sub> forms containing the 16 amino acid sequence [X] produced in accordance with the previously described methods are transferred to nitrocellulose membranes and tested for their ability to bind <sup>32</sup>P-labelled phosphorylase  $\alpha$  in the presence and absence of a test compound. A decrease in the amount of <sup>32</sup>P phosphorylase  $\alpha$  binding in the presence of the test compound compared to the amount of binding in the absence of the test compound is indicative of an inhibitory compound of the invention.

A compound which is capable of blocking the phosphorylase  $\alpha$  G<sub>L</sub> interaction should increase the activity of PP1-G<sub>L</sub> and consequently that of glycogen synthase, leading to the increased conversion of the glucose into glycogen. Hence, the compound should be an effective drug in lowering blood glucose by converting it into hepatic glycogen.

Previously, it was reasonable to assume that since phosphorylase  $\alpha$  is a large molecule, it might bind to many sites or an extensive region of G<sub>L</sub>. Consequently, the probability of finding a drug which could block this interaction was thought to be low. The unexpected identification of the binding site on G<sub>L</sub> has been only 16 amino acids long (or smaller) increases the likelihood of finding a drug that will prevent phosphorylase  $\alpha$  inhibition of the PP1-G<sub>L</sub> complex and hence increase glycogen synthesis.

## 7. *Use in medicine*

The aforementioned compounds of the invention or a formulation thereof may be administered by any conventional method including oral and parenteral (eg subcutaneous or intramuscular) injection. The treatment may consist of a single dose or a plurality of doses over a period of time.

## 8. *Pharmaceutical compositions of the invention*

The following examples illustrate pharmaceutical formulations according to the invention in which the active ingredient is a compound of the invention.

### Example A: Tablet

Active ingredient	100 mg
Lactose	200 mg
Starch	50 mg
Polyvinylpyrrolidone	5 mg
Magnesium stearate	4 mg
	359 mg

Tablets are prepared from the foregoing ingredients by wet granulation followed by compression.

### Example B: Tablet Formulations

The following formulations A and B are prepared by wet granulation of the ingredients with a solution of povidone, followed by addition of magnesium stearate and compression.

Formulation A

	<u>mg/tablet</u>	<u>mg/tablet</u>
(a) Active ingredient	250	250
(b) Lactose B.P.	210	26
(c) Povidone B.P.	15	9
(d) Sodium Starch Glycolate	20	12
(e) Magnesium Stearate	5	3
	—	
	500	300

Formulation B

	<u>mg/tablet</u>	<u>mg/tablet</u>
(a) Active ingredient	250	250
(b) Lactose	150	-
(c) Avicel PH 101 <sup>®</sup>	60	26
(d) Povidone B.P.	15	9
(e) Sodium Starch Glycolate	20	12
(f) Magnesium Stearate	5	3
	—	
	500	300

25

Formulation C

	<u>mg/tablet</u>
Active ingredient	100
Lactose	200
Starch	50
Povidone	5
Magnesium stearate	4

359

The following formulations, D and E, are prepared by direct compression of the admixed ingredients. The lactose used in formulation E is of the direction compression type.

Formulation D

	<u>mg/capsule</u>
Active Ingredient	250
Pregelatinised Starch NF15	150

400

Formulation E

	<u>mg/capsule</u>
Active Ingredient	250
Lactose	150
Avicel <sup>®</sup>	100

500



Formulation F (Controlled Release Formulation)

The formulation is prepared by wet granulation of the ingredients (below) with a solution of povidone followed by the addition of magnesium stearate and compression.

	<u>mg/tablet</u>
(a) Active Ingredient	500
(b) Hydroxypropylmethylcellulose (Methocel K4M Premium) <sup>®</sup>	112
(c) Lactose B.P.	53
(d) Povidone B.P.C.	28
(e) Magnesium Stearate	7
	700

Drug release takes place over a period of about 6-8 hours and was complete after 12 hours.

Example D: Capsule FormulationsFormulation A

A capsule formulation is prepared by admixing the ingredients of Formulation D in Example C above and filling into a two-part hard gelatin capsule. Formulation B (*infra*) is prepared in a similar manner.

Formulation B

	<u>mg/capsule</u>
(a) Active ingredient	250
(b) Lactose B.P.	143
(c) Sodium Starch Glycolate	25
(d) Magnesium Stearate	2
	420

Formulation C

	<u>mg/capsule</u>
(a) Active ingredient	250
(b) Macrogol 4000 BP	350
	600

Capsules are prepared by melting the Macrogol 4000 BP, dispersing the active ingredient in the melt and filling the melt into a two-part hard gelatin capsule.

Formulation D

	<u>mg/capsule</u>
Active ingredient	250
Lecithin	100
Arachis Oil	100
	450

Capsules are prepared by dispersing the active ingredient in the lecithin and arachis oil and filling the dispersion into soft, elastic gelatin capsules.

#### Formulation E (Controlled Release Capsule)

The following controlled release capsule formulation is prepared by extruding ingredients a, b, and c using an extruder, followed by spheronisation of the extrudate and drying. The dried pellets are then coated with release-controlling membrane (d) and filled into a two-piece, hard gelatin capsule.

	<u>mg/capsule</u>
(a) Active ingredient	250
(b) Microcrystalline Cellulose	125
(c) Lactose BP	125
(d) Ethyl Cellulose	13
	513

#### Example E: Injectable Formulation

<u>Active ingredient</u>	0.200 g
--------------------------	---------

Sterile, pyrogen free phosphate buffer (pH7.0) to 10 ml

The active ingredient is dissolved in most of the phosphate buffer (35-40°C), then made up to volume and filtered through a sterile micropore filter into a sterile 10 ml amber glass vial (type 1) and sealed with sterile closures and overseals.

Example F: Intramuscular injection

Active ingredient	0.20 g
Benzyl Alcohol	0.10 g
Glucofurol 75°	1.45 g
Water for Injection q.s. to	3.00 ml

The active ingredient is dissolved in the glycofurol. The benzyl alcohol is then added and dissolved, and water added to 3 ml. The mixture is then filtered through a sterile micropore filter and sealed in sterile 3 ml glass vials (type 1).

Example G: Syrup Suspension

Active ingredient	0.2500 g
Sorbitol Solution	1.5000 g
Glycerol	2.0000 g
Dispersible Cellulose	0.0750 g
Sodium Benzoate	0.0050 g
Flavour, Peach 17.42.3169	0.0125 ml
Purified Water q.s. to	5.0000 ml

The sodium benzoate is dissolved in a portion of the purified water and the sorbitol solution added. The active ingredient is added and dispersed. In the glycerol is dispersed the thickener (dispersible cellulose). The two dispersions are mixed and made up to the required volume with the purified water. Further thickening is achieved as required by extra shearing of the suspension.

Example H: Suppository

	<u>mg/suppository</u>
Active ingredient (63 $\mu$ m)*	250
Hard Fat, BP (Witepsol H15 - Dynamit Nobel)	1770
	2020

\*The active ingredient is used as a powder wherein at least 90% of the particles are of 63  $\mu$ m diameter or less.

One fifth of the Witepsol H15 is melted in a steam-jacketed pan at 45°C maximum. The active ingredient is sifted through a 200  $\mu$ m sieve and added to the molten base with mixing, using a silverson fitted with a cutting head, until a smooth dispersion is achieved. Maintaining the mixture at 45°C, the remaining Witepsol H15 is added to the suspension and stirred to ensure a homogenous mix. The entire suspension is passed through a 250  $\mu$ m stainless steel screen and, with continuous stirring, is allowed to cool to 40°C. At a temperature of 38°C to 40°C 2.02 g of the mixture is filled into suitable plastic moulds. The suppositories are allowed to cool to room temperature.

Example I: Pessaries

	<u>mg/pessary</u>
Active ingredient	250
Anhydrate Dextrose	380
Potato Starch	363
Magnesium Stearate	7
	1000

The above ingredients are mixed directly and pessaries prepared by direct compression of the resulting mixture.

The formulations may conveniently be presented in unit dosage form and may be prepared by any of the methods well known in the art of pharmacy. Such methods include the step of bringing into association the active ingredient (compound of the invention) with the carrier which constitutes one or more accessory ingredients. In general the formulations are prepared by uniformly and intimately bringing into association the active ingredient with liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

Formulations in accordance with the present invention suitable for oral administration may be presented as discrete units such as capsules, cachets or tablets, each containing a predetermined amount of the active ingredient; as a powder or granules; as a solution or a suspension in an aqueous liquid or a non-aqueous liquid; or as an oil-in-water liquid emulsion or a water-in-oil liquid emulsion. The active ingredient may also be presented as a bolus, electuary or paste.

A tablet may be made by compression or moulding, optionally with one or more accessory ingredients. Compressed tablets may be prepared by compressing in a suitable machine the active ingredient in a free-flowing form such as a powder or granules, optionally mixed with a binder (eg povidone, gelatin, hydroxypropylmethyl cellulose), lubricant, inert diluent, preservative, disintegrant (eg sodium starch glycolate, cross-linked povidone, cross-linked sodium carboxymethyl cellulose), surface-active or dispersing agent. Moulded tablets may be made by moulding in a suitable machine a mixture of the powdered compound moistened with an inert liquid diluent. The tablets may optionally be coated or scored and may be formulated so as to provide slow or controlled release of the active

ingredient therein using, for example, hydroxypropylmethylcellulose in varying proportions to provide desired release profile.

Formulations suitable for topical administration in the mouth include lozenges comprising the active ingredient in a flavoured basis, usually sucrose and acacia or tragacanth; pastilles comprising the active ingredient in an inert basis such as gelatin and glycerin, or sucrose and acacia; and mouth-washes comprising the active ingredient in a suitable liquid carrier.

Formulations suitable for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The formulations may be presented in unit-dose or multi-dose containers, for example sealed ampoules and vials, and may be stored in a freeze-dried (lyophilised) condition requiring only the addition of the sterile liquid carrier, for example water for injections, immediately prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules and tablets of the kind previously described.

Preferred unit dosage formulations are those containing a daily dose or unit, daily sub-dose or an appropriate fraction thereof, of an active ingredient.

It should be understood that in addition to the ingredients particularly mentioned above the formulations of this invention may include other agents conventional in the art having regard to the type of formulation in question,



Whilst it is possible for a compound of the invention to be administered alone, it is preferable to present it as a pharmaceutical formulation, together with one or more acceptable carriers. The carrier(s) must be "acceptable" in the sense of being compatible with the compound of the invention and not deleterious to the recipients thereof. Typically, the carriers will be water or saline which will be sterile and pyrogen free.

Claims

1. Use in medicine of a compound which is capable of blocking the interaction of phosphorylase *a* with the glycogen-targeting subunit ( $G_L$ ) of protein phosphatase 1.
2. Use as claimed in Claim 1 wherein the compound is for use in the manufacture of a medicament for use in the treatment of a disorder associated with higher than normal blood glucose levels.
3. Use as claimed in Claim 2 wherein the disorder is selected from type I and/or type II diabetes.
4. Use in medicine as claimed in Claim 1 wherein the compound is a polypeptide comprising the 16 mer amino acid sequence PEWPSYLGYEKLYPYY, or a fragment or variant thereof which is capable of binding phosphorylase *a*.
5. Use as claimed in Claim 4 wherein the polypeptide increases the activity of hepatic glycogen synthase.
6. A pharmaceutical composition comprising an inhibitor compound which is capable of blocking the interaction of phosphorylase *a* with the glycogen - targeting subunit ( $G_L$ ) of protein phosphatase 1, together with a pharmaceutically acceptable excipient or carrier.
7. A pharmaceutical composition as claimed in Claim 6 wherein the inhibitor compound comprises a polypeptide having the 16 mer amino acid

sequence PEWPSYLGYEKLYPYY, or a fragment or variant thereof which is capable of binding phosphorylase *a*.

8. A pharmaceutical composition as claimed in Claim 6 wherein the polypeptide consists of a truncated version of the glycogen-targeting subunit of protein phosphatase 1.

9. A method of identifying an inhibitor compound which is capable of blocking the interaction of phosphorylase *a* with the glycogen-targeting subunit of protein phosphatase 1 comprising;

providing a polypeptide comprising the 16 mer amino acid sequence PEWPSYLGYEKLYPYY, or fragment or variant thereof which binds phosphorylase *a*;

providing a test compound; and

comparing the binding of the polypeptide by phosphorylase *a* in the presence or absence of the test compound; an inhibitor being identified by reduced binding in the presence of the test compound.

10. A method as claimed in Claim 9 wherein the phosphorylase *a* is labelled and the binding of phosphorylase *a* to the polypeptide is determined by measuring the amount of label.

11. A method as claimed in Claim 10 wherein phosphorylase *a* is labelled with a label selected from digoxigenin and <sup>32</sup>P or <sup>33</sup>P.

12. A compound which is identifiable by the method of any one of Claims 9 to 11.

13. A method of reducing the blood glucose level of a mammalian animal comprising administering a therapeutically effective amount of an inhibitor compound as defined in any one of Claims 1 to 12.

14. A method as claimed in Claim 13 wherein the mammalian animal is a human.

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>7</sup> :</b> <b>C07K 14/47, A61K 38/17, G01N 33/68</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 00/12549</b> <b>(43) International Publication Date:</b> 9 March 2000 (09.03.00)
<b>(21) International Application Number:</b> PCT/GB99/02761 <b>(22) International Filing Date:</b> 19 August 1999 (19.08.99)  <b>(30) Priority Data:</b> 9818650.5                      27 August 1998 (27.08.98)                      GB  <b>(71) Applicant (for all designated States except US):</b> MEDICAL RESEARCH COUNCIL [GB/GB]; 20 Park Crescent, London W1N 4AL (GB).  <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> COHEN, Patricia, Townsend, Wade [GB/GB]; Inverbay II, Invergowrie DD2 5DQ (GB). ARMSTRONG, Christopher, George [GB/GB]; 29 Lytton Street, Lytton Grove, Dundee DD1 2EU (GB). DOHERTY, Martin, John [GB/GB]; 3rd floor, 14 Steels Place, Morningside, Edinburgh EH10 4QS (GB).  <b>(74) Agent:</b> MILES, John; Eric Potter Clarkson, Park View House, 58 The Ropewalk, Nottingham NG1 5DD (GB).		<b>(81) Designated States:</b> AU, CA, JP, US, European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).  <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> PROTEIN DOMAINS IN THE HEPATIC GLYCOGEN-TARGETTING SUBUNIT OF PROTEIN PHOSPHATASE 1 AND METHODS OF MAKING AND USING THE SAME  <b>(57) Abstract</b>  The present invention relates to compounds useful for treating disorders associated with abnormal blood glucose levels in mammals, such as Type I and Type II diabetes. The invention also relates to methods of making and using such compounds.		

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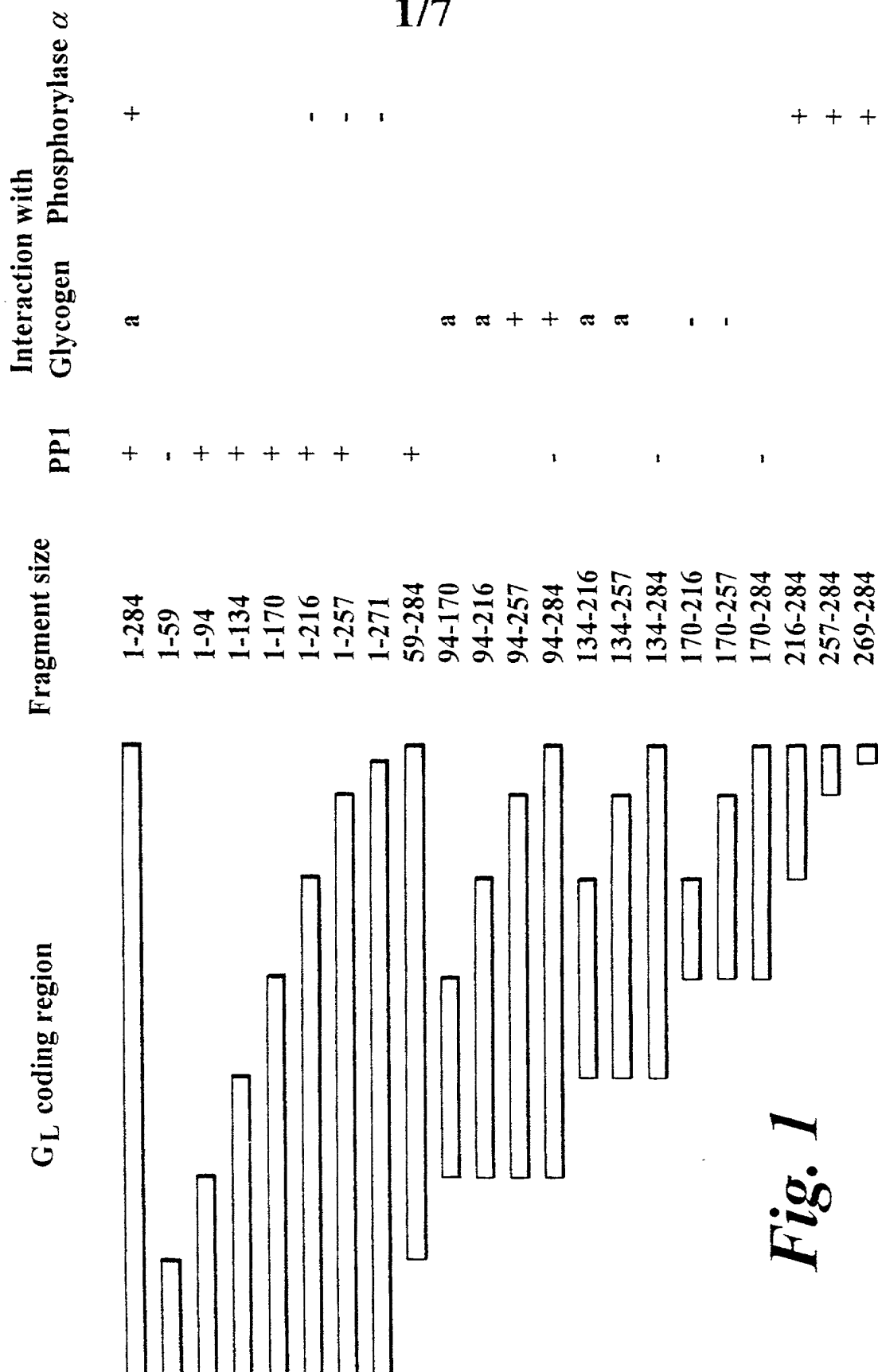
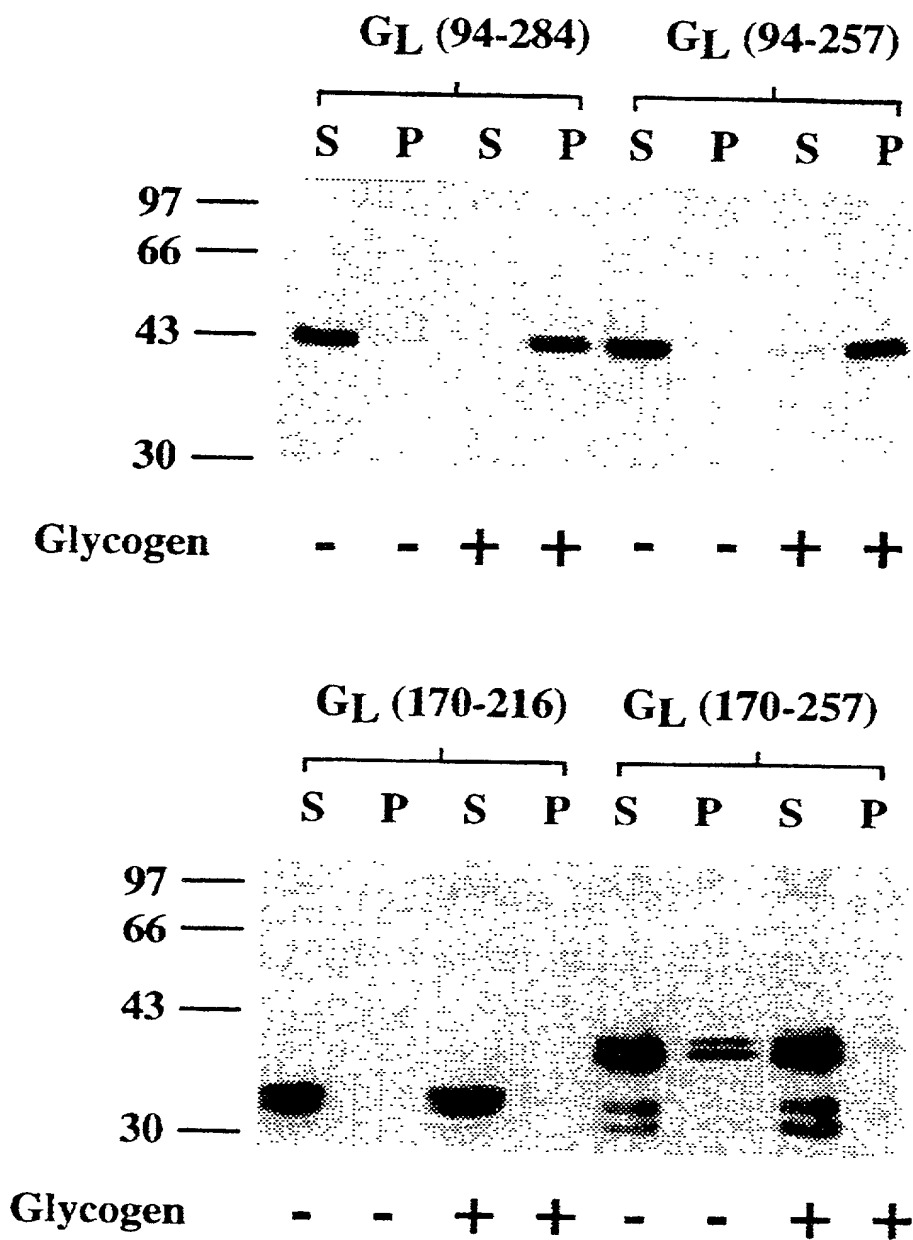
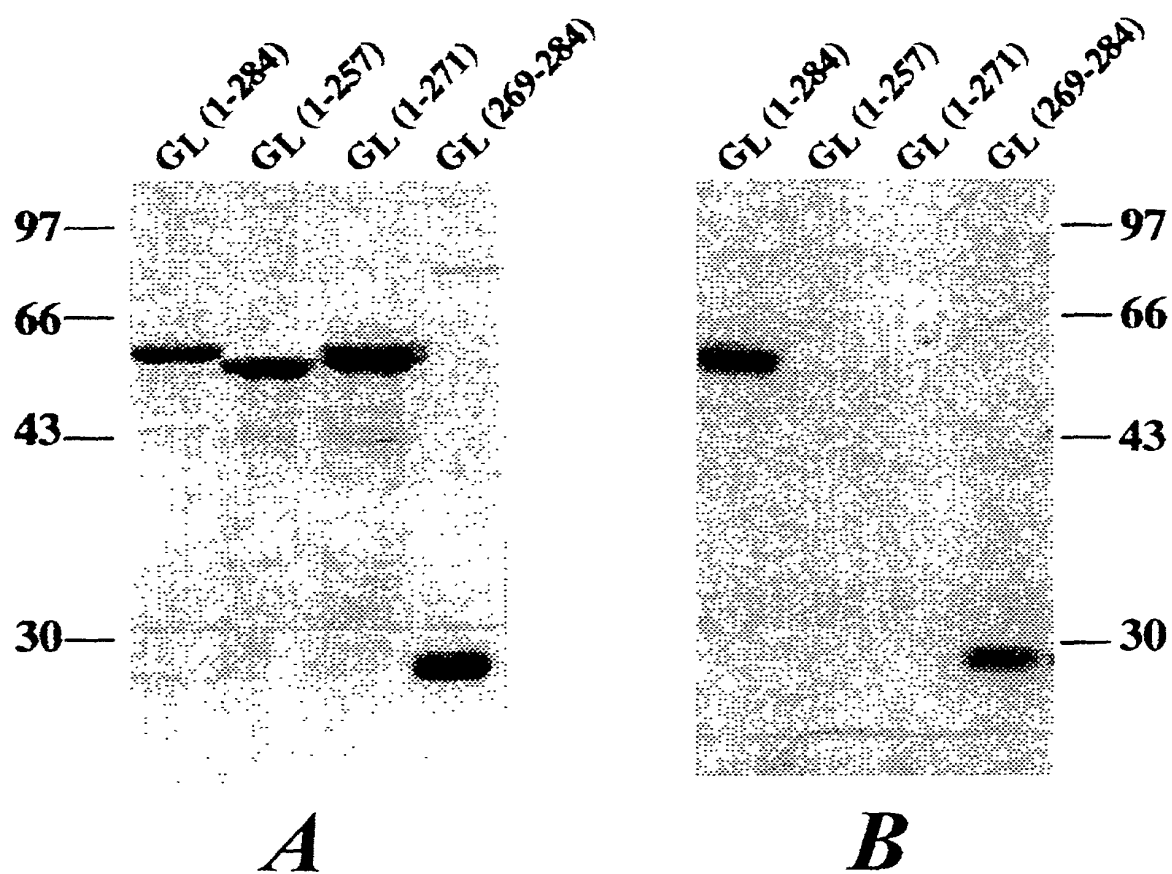


Fig. 1

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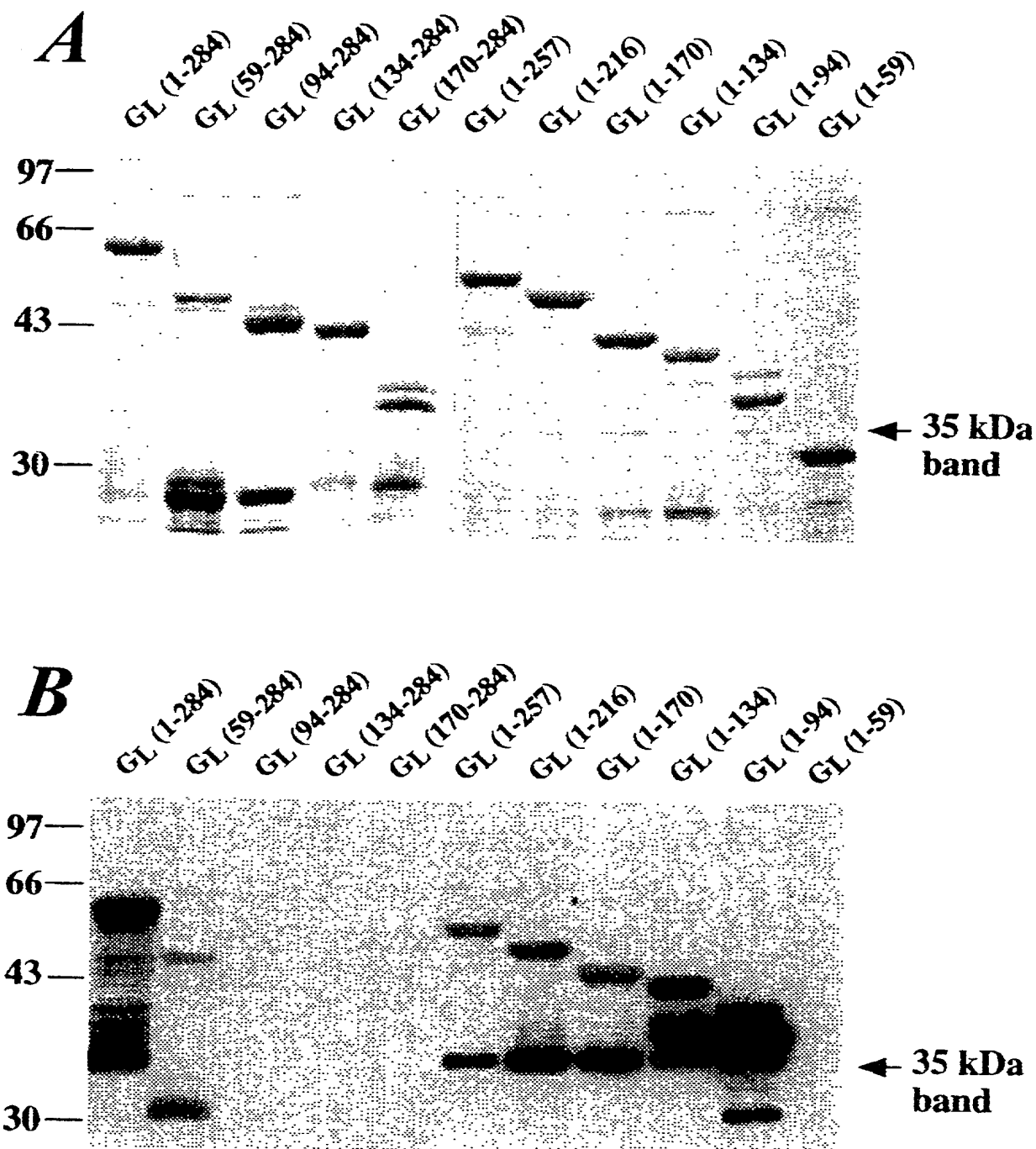
*Fig. 2*

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*Fig. 3*



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**Fig. 4**

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M <sub>110</sub> (1-38)peptide	26	...PPVV <u>K</u> <u>R</u> <u>Q</u> <u>K</u> <u>T</u> <u>K</u> <u>V</u> <u>K</u> <u>F</u>	38
G <sub>L</sub>	52	...TVQ <u>E</u> <u>K</u> <u>K</u> <u>V</u> <u>K</u> <u>R</u> <u>V</u> <u>S</u> <u>F</u> <u>A</u> <u>D</u> <u>Q</u> <u>G</u> <u>L</u> ...	69
G <sub>M</sub> (63-93)peptide	63	G <u>R</u> <u>R</u> <u>V</u> <u>S</u> <u>F</u> <u>A</u> <u>D</u> <u>N</u> <u>E</u> <u>G</u> ...	73

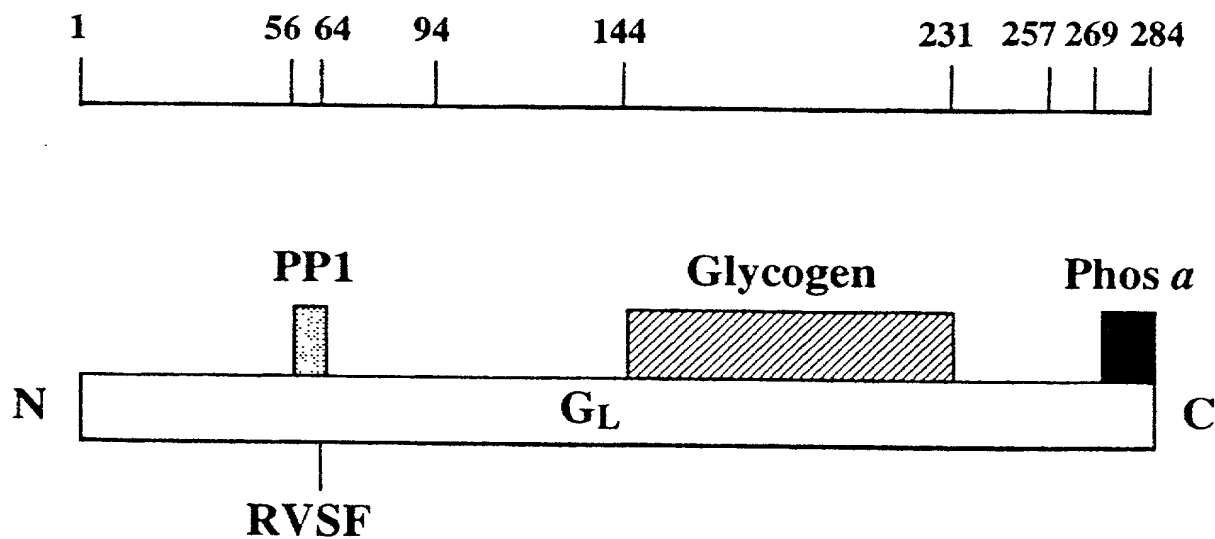
*Fig. 5*

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G <sub>L</sub>	132	VCLE---NCVLKEKAIAGTVKVNLAFEKVVKIRM--TFDTWKSET	172
R5	157	VCLE---NCSLQERTVTGTVKVKNVSEFKVQIRI--TFDSWKNYT	197
R6	177	VCLE---RVTCSDLGISGTVRVCNVAFEKQAVRY--TFSGWRSTH	217
G <sub>M</sub>	128	AI <sub>L</sub> ES- <u>TESLLG</u> STS <u>IKGI</u> IRVLNVSEFKLVYVRM--SLDDWQTHY	170
GAC1	244	VKLHSLTQLGDDSSKITGLVYVKNLSFEKYLEIKF--TFNSWRDIH	287
AMYL	33	VQLDS---YNYDGSTFSGKIYVKNIAYSKKVTVIYADGSDNWNNG	75
Consensus		φ . G . V . V . N φ . F . K . V . V . φ . . . . . W	
		I Y L I	
PHOS	398	RHLQIIYEINQRF <sub>L</sub> LNRVAAAFPGDVDR <sub>L</sub> LRMS	429
		** *** *	
G <sub>L</sub>	173	DFPCQYVKDITYAGSDRD <u>TFSE</u> DISLPEKIQSYE-----	205
R5	198	DVDCVYMKNVYGGTDS <u>TFSE</u> FAIDLPPVIPTEQ-----	230
R6	218	EAVARWRGPAGPEGTE <u>DVF</u> TFGFPVPPFLLELGS-----	251
G <sub>M</sub>	171	DILAEYVPNSCDG-ET <u>DQ</u> SEFKIVLVPPYQKDG-----	203
GAC1	288	YVTANFNRTIN--SNV <u>DE</u> FKFTID <sub>L</sub> NSLKYILLIKRIITMEKNTSS	331
AMYL	76	NTIAASYSAPISGSNYEYWTESASINGIK-----	104
Consensus		D . F . F . . . φ	
		E W	
PHOS	330	LVEEGAVK	437
		* * *	
G <sub>L</sub>	206	---RMEFAVCYECNGQSYWDSNKGKNYRI	231
R5	231	---KIEFCISYHANGQVFDNNDGQNYRI	256
R6	252	---RVHFAVRYQVAGAEYWDNNDHRDYSL	282
G <sub>M</sub>	204	---KVEFCIRYETSVGTFWNNNGTNYTF	229
GAC1	332	CPLNIELCCRYDVNNETYYDNNNGKNYHL	360
AMYL	105	-----EFYIKYEVSGKTYDNNNSANYQV	128
Consensus		F . φ . Y . . . . . YW . . N . . . . Y . φ	
		L FY	

Fig. 6

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*Fig. 7*

# Declaration, Power of Attorney



26474

PATENT & TRADEMARK OFFICE

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0050/050763

We (I), the undersigned inventor(s), hereby declare(s) that:

My residence, post office address and citizenship are as stated below next to my name,

We (I) believe that we are (I am) the original, first, and joint (sole) inventor(s) of the subject matter which is claimed and for which a patent is sought on the invention entitled

Solid Composition Containing Bonds Which Can Be Activated With Actinic Radiation,  
and Its Use

the specification of which

☒ is attached hereto.

☐ was filed on \_\_\_\_\_ as

Application Serial No. \_\_\_\_\_

and amended on \_\_\_\_\_.

☒ was filed as PCT international application

Number PCT/EP00/09626

on February 10, 2000

and was amended under PCT Article 19

on \_\_\_\_\_ (if applicable).

We (I) hereby state that we (I) have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

We (I) acknowledge the duty to disclose information known to be material to the patentability of this application as defined in Section 1.56 of Title 37 Code of Federal Regulations.

We (I) hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed. Prior Foreign Application(s)

Application No.	Country	Day/Month/Year	Priority Claimed
19947521.0	Germany	02 October 1999	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No

We (I) hereby claim the benefit under Title 35, United States Codes, § 119(e) of any United States provisional application(s) listed below.

_____	_____
(Application Number)	(Filing Date)
_____	_____
(Application Number)	(Filing Date)

We (I) hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

Application Serial No.	Filing Date	Status (pending, patented, abandoned)
------------------------	-------------	--

_____	_____	_____
_____	_____	_____
_____	_____	_____
_____	_____	_____

And we (I) hereby appoint **Messrs. HERBERT. B. KEIL**, Registration Number 18,967; and **RUSSEL E. WEINKAUF**, Registration Number 18,495; the address of both being Messrs. Keil & Weinkauff, 1101 Connecticut Ave., N.W., Washington, D.C. 20036 (telephone 202-659-0100), our attorneys, with full power of substitution and revocation, to prosecute this application, to make alterations and amendments therein, to sign the drawings, to receive the patent, and to transact all business in the Patent Office connected therewith.

We (I) declare that all statements made herein of our (my) own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Declaration

Page 3 of 4

0050/050763

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**Declaration**

Page 4 of 4

0050/050763

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Docket No.  
002.00140

# Declaration and Power of Attorney For Patent Application

## English Language Declaration

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name,

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

**PROTEIN DOMAINS IN THE HEPATIC GLYCOGEN-TARGETTING SUBUNIT OF PROTEIN PHOSPHATASE 1 AND METHODS OF MAKING AND USING THE SAME**

the specification of which

(check one)

☐ is attached hereto.

☒ was filed on February 27, 2001 as United States Application No. or PCT International Application Number 09/763,848 and was amended on \_\_\_\_\_

(if applicable)

☐ hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

☐ I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, Code of Federal Regulations, Section 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code, Section 119(a)-(d) or Section 365(b) of any foreign application(s) for patent or inventor's certificate, or Section 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or PCT International application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Priority Not Claimed

9818650.5

GB

27 August 1998

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

(Number)

(Country)

(Day/Month/Year Filed)

☐

I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

\_\_\_\_\_  
(Application Serial No.)

\_\_\_\_\_  
(Filing Date)

\_\_\_\_\_  
(Application Serial No.)

\_\_\_\_\_  
(Filing Date)

\_\_\_\_\_  
(Application Serial No.)

\_\_\_\_\_  
(Filing Date)

I hereby claim the benefit under 35 U. S. C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, C. F. R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

**PCT/GB99/02761**  
\_\_\_\_\_  
(Application Serial No.)

**19 August 1999**  
\_\_\_\_\_  
(Filing Date)

**published**  
\_\_\_\_\_  
(Status)

(patented, pending, abandoned)

\_\_\_\_\_  
(Application Serial No.)

\_\_\_\_\_  
(Filing Date)

\_\_\_\_\_  
(Status)  
(patented, pending, abandoned)

\_\_\_\_\_  
(Application Serial No.)

\_\_\_\_\_  
(Filing Date)

\_\_\_\_\_  
(Status)  
(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (list name and registration number)

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Peter Rogalskyj, Reg. No. 38,601

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**Martin John Doherty**

Third inventor's signature

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Date

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Full name of fourth inventor, if any

Fourth inventor's signature

Date

Residence

Citizenship

Post Office Address

Full name of fifth inventor, if any

Fifth inventor's signature

Date

Residence

Citizenship

Post Office Address

Full name of sixth inventor, if any

Sixth inventor's signature

Date

Residence

Citizenship

Post Office Address